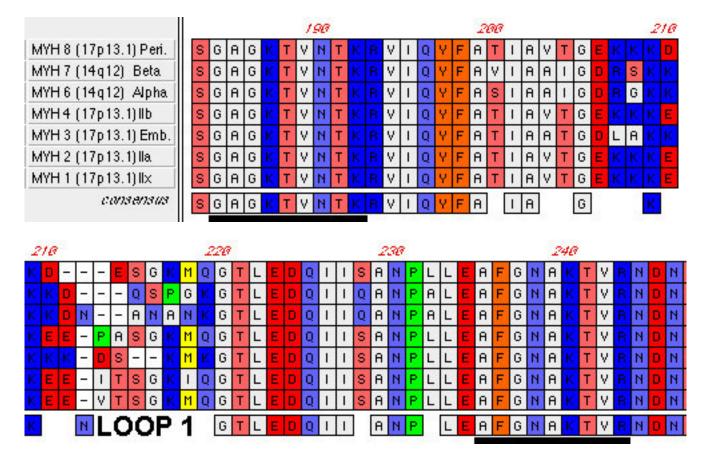
(1) Design of a degenerate PCR primer pair to amplify signature sequences for all vertebrate sarcomeric myosin heavy chain genes : Recognition of the MYH 16 Pseudogene

Based on an alignment of the first seven human sarcomeric myosin heavy chains sequenced, we developed primer pairs to target conserved coding regions in the head domain (bold underline).



The primer pair below flanks the coding region for the hypervariable domain at loop 1 between the 25 kilodalton and the 50 kilodalton proteolytic subfragments of the myosin head. Subcloning of the PCR products was facilitated by the introduction of Sac II and Kpn I sites, as illustrated. Size fractionation of the amplification products was possible on the basis of differences in the length of introns disrupting the coding sequence in the loop 1 domain.

G A GGN GC 5'-CGCC GC	N GGN AA	R CAN GT	N T N AAY CAN I AAY ACI	I AAR	Sense primer
-Sac					
	GGN AAY	GCN AAR	T V CAN GTN TGC CAT Kpn I	NGN GGCG-5 '	Antisense primer

The DNA sequence for a unique 1.1 kb amplification product exactly matched bp 21209-21318 (autoradiograph shown) and 12211-12310 (autoradiograph not shown) of accession number AC112711 (arrow below denotes 3'-end of sense primer).

Α G C T G А G Κ T V N Т K Sense Primer -> CGCCGCGGGIAARACIGTIAAYACIAA... -> 30 20 40 50 60 70 80 10 V ΙQ Y F A N I G G T G K Q T T D K K Κ GAAGGTCATCCAGTACTTTGCCAACATTGGAGGAACTGGCAAACAGACCACAGATAAGAAGqtagagccgaccgggtgggccc GAAGGTCATCCAGTACTTTGCCAACATTGGAGGAACTGGCAAACAGACCACAGATAAGAAGqtagagccgaccgggtgggccc K V I Q Y F A N I G G T G K Q T T D K K 21240 21220 21230 21250 21260 21270 21280 21290 10 90 100 110 intron 7 approximately 1000 bp atatttcccgtcttcaggcttctgtga... atatttcccgtcttcaggcttctgtga... 21300 21310 21320 10 20 30 40 50 60 70 80 DQVI G S L E G S L E D Q V I 22250 22260 22270 22280 22240 22290 22300 22310 90 100 ANPVLE> \bigcirc AGGCAAACCCTGTGCTGGAG... AGGCAAACCCTGTGCTGGAG... A N P V L E> 0 22330 22320 А F G Ν Α Κ Т V R Antisense primer GCN TTY GGN AAY GCN AAR CAN GTN NGN <-3'-CGI AAR CCI TTR CGI TTY TGC CAT GGCG-5'

(2) Other PCR Primers and annealing temperatures

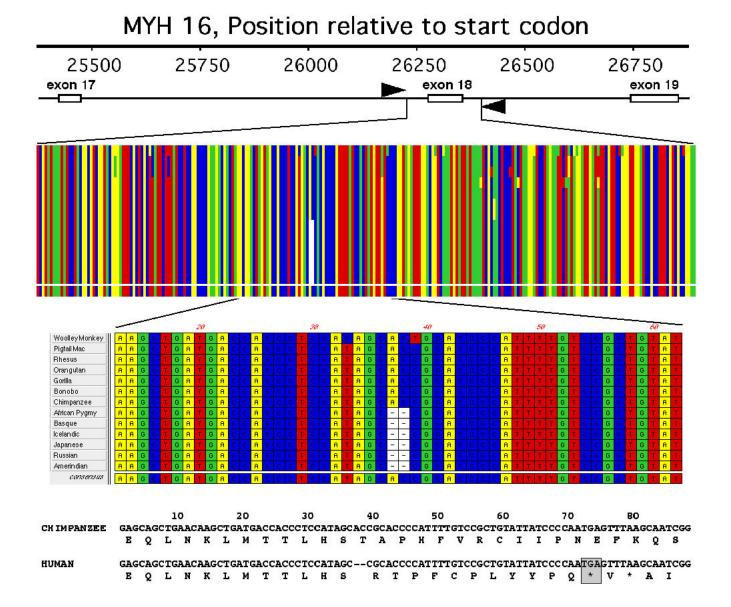
MYH16 RTPCR (Tm=57°C) forward CGGCTCAATCAAATCCTACGG, reverse CTGGCATCCTCGTCCATCTG; *MYH* RTPCR positive control (Tm=47°C) forward GAGGCAAAAAGCGCCAGGA, reverse TTGGTGAAGTTGATGCACAGCT. Genomic DNA Exon 18 (Tm=52°C) forward GTTGAGGTTTCTCTCAGAGCCTTG, reverse CATGTGGGTGCTCTGCAACATC; Exon 23 (Tm=54°C) forward TCTTGGTGTGTGTGTCTTTGC, reverse TAGGGGGCTTTAGGTATGG; Exon 25 (Tm=48°C) forward GGTTCCTTCTTGCCTTCTGAC, reverse TGCCCACCAGGTAATGTGTG; Exon 30 (Tm=48°C) forward CCGATGGTTTGGATTGTG, reverse TAAAGCAGCCTGTGAACG; Exon 31 (Tm=48°C) forward GCTG-TGGCTGTCCTGTAAAC, reverse TCTTCCTGATGACCCCAGAC; Exon 34b (Tm=48°C) forward TGTGACGGTTTTTCCATTAC, reverse TGAGTTAGCCCCCTTTAGG; Exon 37 (Tm=53°C) forward TAGAGCACCCTTTCCACCAAAC, reverse AAGACAGGCATCTCACACACATACC.

(3) Annotation of genomic DNA sequence for the MYH 16 Pseudogene

The *MYH16* locus in the human genome has been annotated as a Third Party Annotation (TPA) and is available under the GenBank TPA accession number: BK001410. Exon 18 sequences from the primate species shown below have been assigned the following GenBank accession numbers:

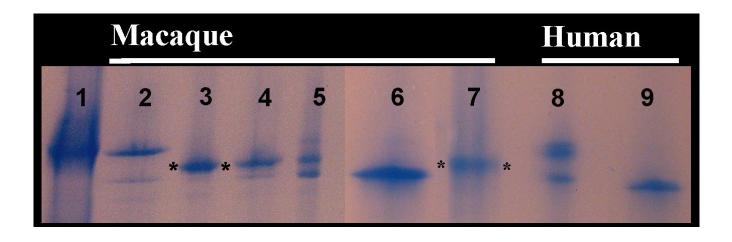
Chimpanzee (Pan troglodytes)	AY350722
Bonobo (Pan paniscus)	AY350721
Gorilla (Gorilla gorilla)	AY350720
Orangutan (Pongo pygmaeus)	AY350719
Pigtailed Macaque (Macaca nemestrina)	AY350717
Rhesus Macaque (Macaca mulatta)	AY350718
Woolley Monkey (Lagothrix lagotricha)	AY350716

(4) Alignment of Human and Non-human Primate DNA sequences for MYH 16 exon 18 and portions of the flanking introns



(5) Myosin gel electrophoresis for LC/MS-MS.

Coomasie blue-stained gel electrophoretograms of myosins from limb and first pharyngeal arch muscles of *M. fascicularis* and *H. sapiens* showing an isomyosin of unique mobility in the first pharyngeal arch derivatives of *M. fascicularis*, midway between fast-IIa/IIx comigrating at top and slow-I at bottom in lane 8. Labeling: 1, m.w. standard; 2, tibialis anterior, 3, temporalis (asterisk denotes band cut for proteomic analysis below); 4, masseter; 5, tensor veli palatini; 6, soleus; 7, temporalis; 8, *H. sapiens* temporalis; 9, *H. sapiens* soleus. Note that the topmost band in lane 8 is a doublet, corresponding to the products of the *MYH 1* and 2 genes¹.



(6) Peptide analysis by LC/MS-MS.

The *M. fascicularis* temporalis myosin heavy chain band of unique electrophoretic mobility was extracted and processed for LC/MS-MS analysis in the Wistar Institute proteomics core as described: http://www.wistar.upenn.edu/research_facilities/facilities/protein/service.html#a

Sequences highlighted in red correspond to positions of 32 peptides identified by LC-MS/MS analysis of a tryptic digest of the predominant myosin in the *M. fascicularis* temporalis muscle (major band in lane e-3). All peptides underlined correspond to confirmed high-probability spectral matches obtained in a SEQUEST search of a current non-redundant protein sequence database updated to include the predicted MYH 14, 15 and 16 gene products. Cross comparison of these to the MYH gene products aligned in the attached file "MYH 1-8 Peptide Alignment" demonstrates the uniqueness of the majority of these peptide sequence motifs.

MPGGYK <mark>GECG</mark>	DDVDPMPFLA	PPEK ERIEAM	NKPYDIKRSC	WVKDEK <mark>EGFV</mark>	AGEIQSEQGD	QVTVKTITNQ	TLTVKKDDIQ
QMNPPKFYQA	SDMADMTFLN	EASVLDNLRQ	RYTNMRIYTY	SGLFCVTVNP	YKWLPIYGAR	VANMYKGKKR	TEMPPHLFSI
SDNAYHDMLM	DRENQSMLIT	GESGAGK TEN	TKKVIQYFAN	IGGTGKQTTD	KKGSLEDQVI	QANPVLEAFG	NAK TTRNNNS
SRFGKFIRIH	FGTTGK <mark>LAGA</mark>	DIESYLLEKS	RVISQQAAER	SYHIFYQILS	NKKPELVESL	LLVPNPKEYH	WVSQGVTTVD
NMDDKEELQI	TDEAFDVLGF	SAEEKMAVYK	LTGGIMHFGN	MKFKQKPR <mark>DE</mark>	QAEVDTTEVA	DKVAHLMGLN	<u>SGELQK</u> GITR
PRVKVGNEFV	QKGQNMEQCQ	NSIGALGKAV	YDKMFKWLVA	RINKTLDTKM	QRQFFIGVLD	IAGFEIFEFN	SFEQLCINFT
NEKLQQFFNH	HMFVLEQEEY	KREGIEWVFI	DFGLDLQACI	DLLEKPMGIF	SILEEQCVFP	KATDATFKAA	LYDNHLGK <mark>SS</mark>
<u>NFLKPK</u> GGKS	KGPEVHFELV	HYAGTVGYNI	TGWLEK <mark>NKDP</mark>	LNETVVGLFQ	<u>K</u> SSVAILALL	FKEEEAPAGS	KKQKR <mark>GSSFM</mark>
TVSNFYREQL	NKLMTTLHST	APHFVRCIIP	NEFKQSGVID	AHLIMHQLAC	NGVLEGIRIC	RKGFPNRLQY	PEFKQR <mark>YQVL</mark>
NPNVIPQGFV	DNKKASELLL	AAIDLDVNEY	KIGHTKVFFR	AGILARLEDM	RDERLAKIMT	MLQCRLRGFL	MRVEFKKMLE
RRMGLKVIQQ	NVHKFLQLRF	WGWWKLYNKV	KPLLNVARQE	EEMKAKEEEL	RKAMAQTQEL	VNKVKELEEK	TATLSQEKND
LTIQLQAEQE	NLMDAEERLT	WMMKTKMDLE	SQISDMRERL	EEEEGMAASL	SAAKRKLEGE	LSDLKRDLEG	LETTLAKTEK
EKQALDHKVR	TLTGDLSLRE	DSITKLQKEK	RALEELHQKT	LDDLQAEEDK	VNHLTKNNSK	LSTQIHELED	NWEQEKKIRA
EVEKARRKAE	SDLKMTIDNL	NEMER <mark>SKLDL</mark>	EEVVK KRDLE	INSVNSK <mark>YED</mark>	EQSLNSTLQR	KLKEHQDR <mark>IE</mark>	ELEEELEAER
AMRAKVEKQR	SDLSR <mark>DLEDL</mark>	SDRLEEAGGA	TSAQIEQNRK	REAELLKLRR	ELEEAALQSE	ATASTLRKKH	VDSMAELTEH
VESLQRVKSK	LEKDKQVMKA	EIDDLNASME	TIQKSKMNAE	AHVR <u>KLEDSL</u>	SEANAK VAEL	ERNQAEINAI	RTRLQAENSE
LSREYEESQS	RLNQILRIK <mark>T</mark>	SLTSQVDDYK	RQLDEESKSR	STAVVSLANT	KHDLDLVKEQ	LEEEQGGKSE	LQRLVSK <mark>LNT</mark>
EVTTWRTKYE	TDAIQRTEEL	EETKRKLAAR	LQEAEEAAET	AQARAASLEK	NKQRLQAEVE	DLTIDLEKAN	AAAAALDKKQ
RLFDK <mark>MLAEW</mark>	QQK CEELQVE	VDSSQKECR <mark>M</mark>	YMTESFK IKT	AYEESLEHLE	SVKKENKTLQ	EEIKDLIDQL	GEGGRSVHEL
QKLKKKLEME	KEELQVALEE	AESSLEVEES	KVIRIQLELA	QVKADIDRR1	HEKEEEFEAT	R KNHQRAIES	LQASLEAEAK
GRAEALRLKK	KMETDLNEME	IQLDHANKNN	SELVKTLKRL	QQQIKDLQVQ	MDEDARQHEE	LRKQYNLQER	RLSLLQTELE
EVR SALEGSE	RSRKLLEQEV	VEITEWHNEI	NIQNQSLLVV	KRKLESDVQR	ISNEHEELIS	EFR LTEERAK	KAMMDAARMA
EELRQEQDHC	MHLEKIKKNY	EVTIKDLQAK	MEEAEQLALK	GGKRTIMKLE	ARIKELETEL	DGEQKQHVET	VKTLCKNERR
LKELVFQTEE	DHKTNQRMQA	LVEKLQNKLK	VYKR <mark>QIEEAE</mark>	DQANQTLARY	RKTVHELDDA	EDRAGMAETA	LNKLRTRHRV
AGK <mark>GITSVEI</mark>	IQVSK TGTSK	TLSEE					

(7) Reconstruction of Ancestral Sequences

The limited availability of fresh masticatory muscle samples from endangered primate species required us to sequence coding regions from genomic DNA. We selected a sampling of the largest exons deduced from our human MYH 16 pseudogene reconstruction and designed PCR primers from the flanking intron sequences. Sequence for Canis familiaris and Macaca fascicularis were obtained by RTPCR of mRNA prepared from biopsies of temporalis muscle. These sequences are available under the following Genbank Accession numbers (corresponding sequence from *Homo sapiens* can be found in our Third Party Annotation of the MYH16 locus, as described above – TPA# BK001410):

Species/Exon	<u>23</u>	<u>25</u>	<u>30</u>	<u>31</u>	<u>34b</u>	<u>37</u>
Chimpanzee (Pan troglodytes)	AY350724	AY350726	AY485943	AY485947	AY350730	AY350728
Orangutan (Pongo pygmaeus)	AY485954	AY485955	AY485956	AY485957	AY485948	AY485958
Cynomolgus Macaque (Macaca fascicularis)	AY350723	AY350725	AY485942	AY485945	AY350729Ź	AY350727
Dog (Canis familiaris)	AY485951	AY485952	AY485944	AY485946	AY485950	AY485953
Gorilla (Gorilla gorilla)					AY485949	

These sequences were compiled into a contiguous open reading frame to facilitate codon-by-codon analysis. An unambiguous alignment of these sequences (no insertions or deletions were required) was obtained with the program ClustalW, as implemented in the MacVector software package (Accelrys, Symantec Corporation). This sequence alignment was exported into the appropriate file format for ancestral sequence reconstruction by two algorithms: maximum parsimony and maximum likelihood. We used the program PAMP for the former and BASEML for the latter, both as contained within the most recently posted release of the PAML software package, <u>paml3.13d.tar.gz</u>, from the ftp download website: <u>ftp://abacus.gene.ucl.ac.uk/pub/paml/</u>

The reconstructed ancestral sequences were identical at all nucleotide positions with the sole exception of an ambiguity which was resolved by sequencing exon 34b of the MYH 16 ortholog from gorilla and rerunning the analysis for this exon (AY485949). Ancestral sequences are available from the author upon request as either merged files or as separate files for the individual exons.

The combined ancestral and extant species sequence file alignment was converted into a file format native to the MEGA2 software package as downloaded from http://www.megasoftware.net/ and used to calculate synonymous and non-synonymous mutations for all pairwise sequence comparisons. In addition, statistics on the probability of negative (purifying) darwinian selection were calculated using the Z-test function as implemented in the MEGA2 package². Relevant data are summarized in Figure 4, a complete spreadsheet is available in the MSExcel file: MYH 16 Distances.xls.

(8) Statistical Significance of Differences in dN/dS Ratios Between Lineages

To supplement the *ad hoc* calculations on statistical significance in the MYH 16 Distances.xls file, we used the method of Yang³ and the software described therein (specifically the codeml program of the PAML package) to examine the statistical significance of the lineage-specific differences in dN/dS ratios (ω) seen in our sampling of the MYH 16 coding sequence. This statistical test allows one to assess whether a two-ratio model fits the data better than a one-ratio model by comparing the likelihood (I) given by maximum likelihood (ML) analysis for each branch when ω is constrained to be the same as all other branches (the null hypothesis), with I given when ω for that branch was free to settle upon its own value. Only in the case of the human lineage does the difference between these ω values achieve a statistical significance of approximately p = 0.05, with a lineage-specific ω of approximately 0.5. Other lineage-specific values of ω average approximately 0.07, with a maximum of 0.12, implying purifying darwinian selection. In all cases an overall tree topology similar to that in Figure 4 was assumed.

Table 1 dN/dS ratios and log likelihood values under different models							
Model	dN/dS Ratios Allowed	Free Parameter	р	1	ω	ω _{free}	
А	1	none	12	1897.49	0.0522	_	
В	2	ω _D	13	1896.50	0.0746	0.0376	
С	2	ω _M	13	1897.38	0.0499	0.0718	
D	2	ω _P	13	1896.85	0.0471	0.1168	
Е	2	$\omega_{\rm C}$	13	1897.36	0.0526	0.0001	
F	2	ω _H	13	1895.62	0.0478	0.5120	

Table 1 dN/dS ratios and log likelihood differences under different models

Model A constraints the dN/dS ratio (ω) to be the same for all branches. Models B-F allow the dN/dS ratio for a single branch (species) to vary from the others. p, number of parameters; l, log likelihood value; ω_0 represents the background dN/dS ratio among all branches not free to vary; ω_D , dog; ω_M , macaque; ω_P , pongo (orang-utan); ω_C , chimpanzee; ω_H , human.

Null Hypothesis Tested	Assumption Made	Models Compared	2ΔΙ	<i>p</i> -value
$\omega_{\rm D} = \omega_0$	$\omega_{\rm M} = \omega_{\rm P} = \omega_{\rm C} = \omega_{\rm H} = \omega_0$	A and B	1.986796	0.1587
$\omega_M = \omega_0$	$\omega_{\rm D} = \omega_{\rm P} = \omega_{\rm C} = \omega_{\rm H} = \omega_0$	A and C	0.229252	0.6320
$\omega_{P} = \omega_0$	$\omega_{\rm D} = \omega_{\rm M} = \omega_{\rm C} = \omega_{\rm H} = \omega_0$	A and D	1.277616	0.2583
$\omega_{C}=\omega_{0}$	$\omega_{\rm D} = \omega_{\rm M} = \omega_{\rm P} = \omega_{\rm H} = \omega_0$	A and E	0.256468	0.6125
$\omega_{\rm H} = \omega_0$	$\omega_{\rm D} = \omega_{\rm M} = \omega_{\rm P} = \omega_{\rm C} = \omega_0$	A and F	3.753482	0.0527*

Table 2 Significance of log likelihood value difference between models

Table 2 Significance of log likelihood value differences between models

For each model described in Table 1, we test the null hypothesis that the dN/dS ratio along each branch is the same whether it is free to vary (Models B-F) or is constrained to have the same dN/dS ratio as all of the other branches (Model A).

(9) Estimation of the age of the MYH 16 gene inactivation

See Chou et al., 2002, Proc Natl Acad Sci USA 11736-41, for a detailed description of the rationale for this analysis. Briefly, the assumption is made that non-synonymous mutations are selected against until the gene is inactivated, thereafter mutations at both synonymous and non-synonymous sites accumulate at the neutral mutation rate. Quantification of lineage-specific mutation rates at synonymous and non-synonymous sites remote from the inactivating deletion provides the information necessary for the calculation. We define the following terms as described in Chou, et al., 2002:

t = time since last common ancestor (LCA) human/chimpanzee

 t_1 = time since MYH 16 deletional inactivation

k = neutral mutation rate = (number synonymous mutations/number of synonymous sites)/t
= dS/t = 1/2 neutral site divergence human-chimpanzee/t

 $f_N = dN/dS$ (average for all species) = .046 from data in MYH 16 Distances.xls and Figure 4.

 $f_N k$ (t-t₁) + kt₁ = dN(LCA to human lineage) LCA is represented by Node C in Figure 4.

 $f_Nkt + kt_1(1-f_N) = dN(LCA \text{ to human lineage})$

 $t_1(1-f_N) = (dN(LCA \text{ to human lineage})/k) - f_N t$

 $t_1 = ((dN(LCA \text{ to human lineage})/k) - f_N t) / (1-f_N)$

= (($dN(LCA \text{ to human})/(dS(LCA \text{ to Human})) - f_N) / (1-f_N)$) t

= ((dN(LCA to human)/(1/2 human-chimp neutral site divergence) - f_N) / (1- f_N)) t

The calculation requires normalization to the fossil record, for which we focus on the humanchimpanzee divergence to minimize the effects of lineage-specific slowing of the molecular clock⁴. Recent fossil data suggests a divergence as remote as 6 to 7 mya⁵. The level of codon bias in the *MYH* genes is high (data not shown), perhaps reflecting their genomic contexts or their high levels of expression⁶. This may diminish the rate of the silent molecular clock as determined from MYH synonymous positions. Thus we substitute the silent site mutation rate obtained from a genome-wide sampling of non-coding, non-repetitive human and chimpanzee DNA sequences for the minimal sampling of non-synomymous sites represented by our sequencing of MYH 16 orthologs⁴. This global number has been defined with precision as the number of substitutions per 100 sites between the two species: 1.19 + -0.016 (Yi). Substituting the foregoing numbers into the last equation above we obtain:

 $t_1 = ((2/840) / ((1/2) * .0119 + 0.00016) - .04643) / (1-.04643)) 6.5 + 1 mya = (.37 + 0.01) (6.5 + 1)$

= 2.4 +/- .25 mya

We recognize that with the current data sampling this estimate is based on a small number of nonsynoymous substitutions between human and chimpanzee, as is necessarily the case. Although this calculation yields a broad temporal range, we note that hominid fossils exhibiting decreased masticatory robusticity fall into the recent end of this window.

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- 4. Yi, S., Ellsworth, D. L. & Li, W. H. Slow molecular clocks in Old World monkeys, apes, and humans. *Mol Biol Evol* **19**, 2191-8 (2002).
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